

An Introduction to Systems Biology from a Machine Learning Perspective II

Neil D. Lawrence

work with Magnus Rattray, Pei Gao, Antti Honkela, Michalis Titsias
and Jennifer Withers

Tampere University of Technology, Finland

23rd June 2009

Roadmap

- 1 GPs and Differential Equations
- 2 Cascaded Differential Equations
- 3 Non-linear Response Models
- 4 Discussion and Future Work
- 5 Acknowledgements

Roadmap

- 1 GPs and Differential Equations
- 2 Cascaded Differential Equations
- 3 Non-linear Response Models
- 4 Discussion and Future Work
- 5 Acknowledgements

Quoting from Khanin *et al.*:

One can come up with linear (or higher order) $f(t)$ approximations on each subinterval. This will introduce additional parameters, which will be impossible to infer with any certainty given limited amount of data.

Khanin *et al.* (2006)

- Linear Activation Model (Barenco et al., 2006, Genome Biology)

$$\frac{dx_j(t)}{dt} = B_j + S_j f(t) - D_j x_j(t)$$

- $x_j(t)$ – concentration of gene j 's mRNA
- $f(t)$ – concentration of active transcription factor
- Model parameters: baseline B_j , sensitivity S_j and decay D_j
- Application: identifying co-regulated genes (targets)
- Problem: how do we fit the model when $f(t)$ is not observed?

- Linear Activation Model (Barenco et al., 2006, Genome Biology)

$$\frac{dx_j(t)}{dt} = B_j + S_j f(t) - D_j x_j(t)$$

- $x_j(t)$ – concentration of gene j 's mRNA
- $f(t)$ – concentration of active transcription factor
- Model parameters: baseline B_j , sensitivity S_j and decay D_j
- Application: identifying co-regulated genes (targets)
- Problem: how do we fit the model when $f(t)$ is not observed?

- Linear Activation Model (Barenco et al., 2006, Genome Biology)

$$\frac{dx_j(t)}{dt} = B_j + S_j f(t) - D_j x_j(t)$$

- $x_j(t)$ – concentration of gene j 's mRNA
- $f(t)$ – concentration of active transcription factor
- Model parameters: baseline B_j , sensitivity S_j and decay D_j
- Application: identifying co-regulated genes (targets)
- Problem: how do we fit the model when $f(t)$ is not observed?

- Linear Activation Model (Barenco et al., 2006, Genome Biology)

$$\frac{dx_j(t)}{dt} = B_j + S_j f(t) - D_j x_j(t)$$

- $x_j(t)$ – concentration of gene j 's mRNA
- $f(t)$ – concentration of active transcription factor
- Model parameters: baseline B_j , sensitivity S_j and decay D_j
- Application: identifying co-regulated genes (targets)
- Problem: how do we fit the model when $f(t)$ is not observed?

- Linear Activation Model (Barenco et al., 2006, Genome Biology)

$$\frac{dx_j(t)}{dt} = B_j + S_j f(t) - D_j x_j(t)$$

- $x_j(t)$ – concentration of gene j 's mRNA
- $f(t)$ – concentration of active transcription factor
- Model parameters: baseline B_j , sensitivity S_j and decay D_j
- Application: identifying co-regulated genes (targets)
- Problem: how do we fit the model when $f(t)$ is not observed?

- Linear Activation Model (Barenco et al., 2006, Genome Biology)

$$\frac{dx_j(t)}{dt} = B_j + S_j f(t) - D_j x_j(t)$$

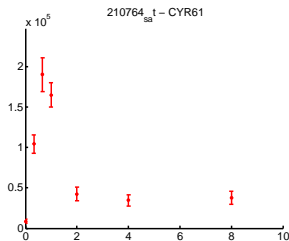
- $x_j(t)$ – concentration of gene j 's mRNA
- $f(t)$ – concentration of active transcription factor
- Model parameters: baseline B_j , sensitivity S_j and decay D_j
- Application: identifying co-regulated genes (targets)
- Problem: how do we fit the model when $f(t)$ is not observed?

Why use a model-based approach?

- Co-regulated genes can differ greatly in their expression profiles

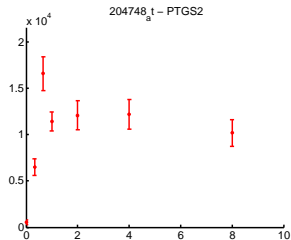
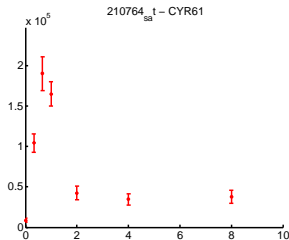
Why use a model-based approach?

- Co-regulated genes can differ greatly in their expression profiles



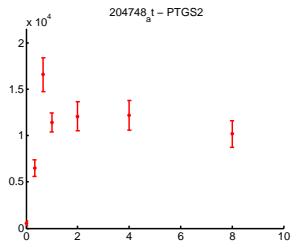
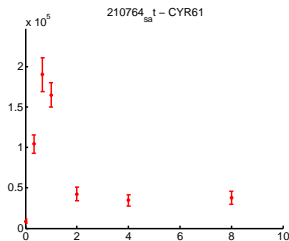
Why use a model-based approach?

- Co-regulated genes can differ greatly in their expression profiles



Why use a model-based approach?

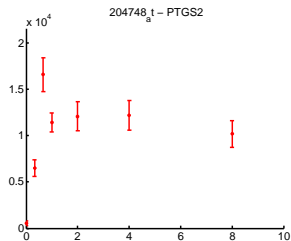
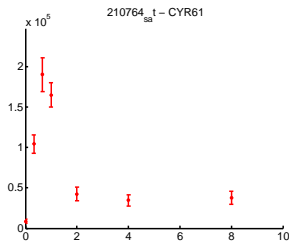
- Co-regulated genes can differ greatly in their expression profiles



- Clustering cannot be relied on to identify co-regulated genes

Why use a model-based approach?

- Co-regulated genes can differ greatly in their expression profiles



- Clustering cannot be relied on to identify co-regulated genes
- A model-based approach is required

Models of non-linear regulation

- Non-linear Activation: Michaelis-Menten Kinetics

$$\frac{dx_j(t)}{dt} = B_j + \frac{S_j f(t)}{\gamma_j + f(t)} - D_j x_j(t)$$

used by Rogers and Girolami (2006)

- Non-linear Repression

$$\frac{dx_j(t)}{dt} = B_j + \frac{S_j}{\gamma_j + f(t)} - D_j x_j(t)$$

used by Khanin et al., 2006, PNAS 103

Models of non-linear regulation

- Non-linear Activation: Michaelis-Menten Kinetics

$$\frac{dx_j(t)}{dt} = B_j + \frac{S_j f(t)}{\gamma_j + f(t)} - D_j x_j(t)$$

used by Rogers and Girolami (2006)

- Non-linear Repression

$$\frac{dx_j(t)}{dt} = B_j + \frac{S_j}{\gamma_j + f(t)} - D_j x_j(t)$$

used by Khanin et al., 2006, PNAS 103

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

Standard inference approach

- Previous approaches all use similar inference methodology:
 - ▶ Represent $f(t)$ as coarse-grained piecewise continuous function $[f_1, f_2, \dots, f_d]$
 - ▶ Often discretize where data are collected
 - ▶ Treat f_i as additional model parameters
 - ▶ Use maximum likelihood or Bayesian MCMC to estimate $\{f_i\}$ along with other model parameters of interest
- Limitations:
 - ▶ Arbitrary choice of discretization points
 - ▶ Coarse-grain gives crude approximation to $f(t)$
 - ▶ Fine-grain leads to harder inference problem

- Gaussian Process

$$f(t) \sim \mathcal{GP}(m(t), k(t, t'))$$

where

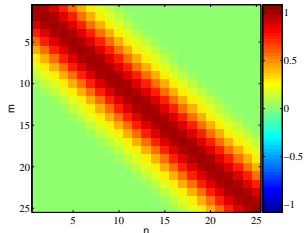
$$\begin{aligned} m(t) &= \mathbb{E}[f(t)] = \langle f(t) \rangle \\ k(t, t') &= \mathbb{E}[(f(t) - m(t))(f(t') - m(t'))] \end{aligned}$$

► Skip Covariance Functions

RBF Kernel Function

$$k(t, t') = \alpha \exp \left(-\frac{(t - t')^2}{2l^2} \right)$$

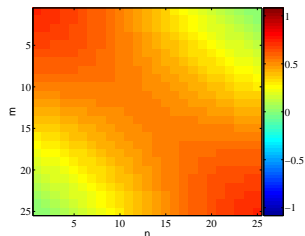
- Covariance matrix is built using the *inputs* to the function t .
- For the example above it was based on Euclidean distance.
- The covariance function is also known as a kernel.



MLP Kernel Function

$$k(t, t') = \alpha \sin^{-1} \left(\frac{w t t' + b}{\sqrt{w t^2 + b + 1} \sqrt{w t'^2 + b + 1}} \right)$$

- A non-stationary covariance matrix (Williams, 1997).
- Derived from a multi-layer perceptron (MLP).



demCovFuncSample

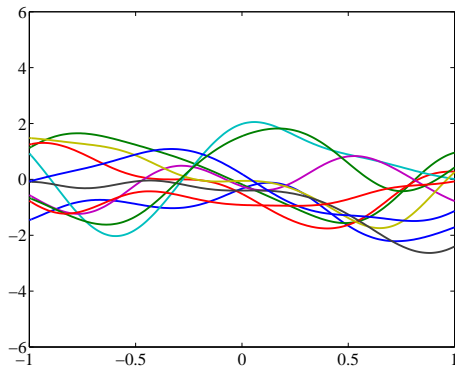


Figure: RBF kernel with $\gamma = 10^{-\frac{1}{2}}$, $\alpha = 1$

demCovFuncSample

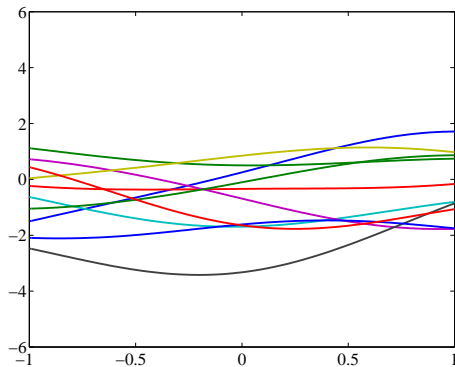


Figure: RBF kernel with $l = 1$, $\alpha = 1$

demCovFuncSample

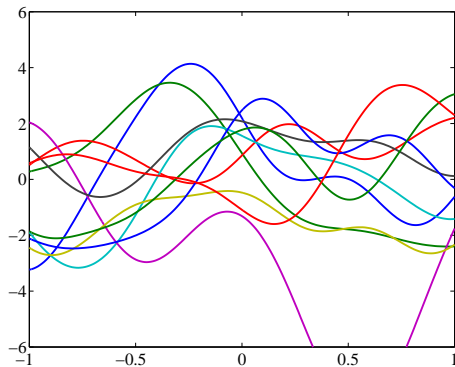


Figure: RBF kernel with $l = 0.3$, $\alpha = 4$

demCovFuncSample

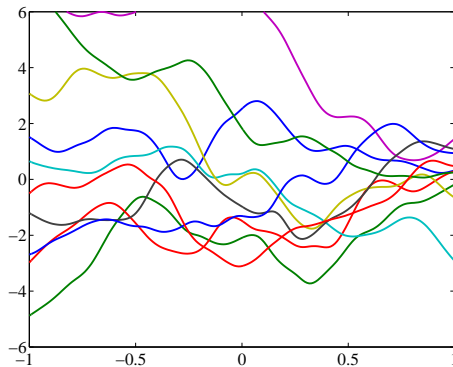


Figure: MLP kernel with $\alpha = 8$, $w = 100$ and $b = 100$

demCovFuncSample

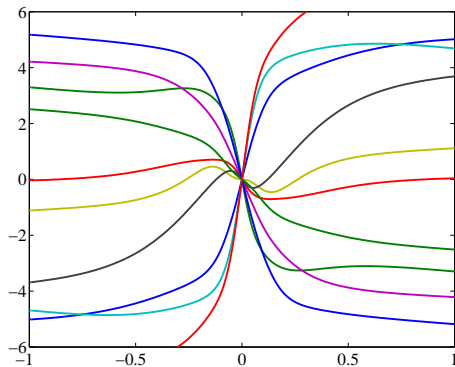


Figure: MLP kernel with $\alpha = 8$, $b = 0$ and $w = 100$

Linear Activation Model

Recall the linear model

$$\frac{dx_j(t)}{dt} = B_j + S_j f(t) - D_j x_j(t) .$$

This differential equation can be solved for $x_j(t)$ as

$$x_j(t) = \frac{B_j}{D_j} + S_j \int_0^t e^{-D_j(t-u)} f(u) du .$$

Note: This is a linear operation on $f(t)$.

If $f(t)$ is a zero mean Gaussian process then $x_i(t)$ is also a Gaussian process with mean $\frac{B_i}{D_i}$.

► Skip GP Properties

Two Properties of GPs

The integral of a GP is also a GP,

$$f(t) \sim N(\mathbf{0}, \mathbf{K}_{ff})$$

and

$$g(t) = \int_0^t f(u) \, du$$

then

$$g(t) \sim N(\mathbf{0}, \mathbf{K}_{gg}),$$

where

$$k_{gg}(t, t') = \int_0^t \int_0^{t'} k_{ff}(u, u') \, du \, du'$$

Two Properties of GPs

Product with deterministic function

Product with a deterministic function leads to another GP,

$$f(t) \sim N(\mathbf{0}, \mathbf{K}_{ff}),$$

and

$$g(t) = f(t) h(t)$$

where $h(t)$ is a deterministic function then,

$$g(t) \sim N(\mathbf{0}, \mathbf{K}_{gg}),$$

where

$$k_{gg}(t, t') = h(t) k_{ff}(t, t') h(t')$$

Covariance for Transcription Model

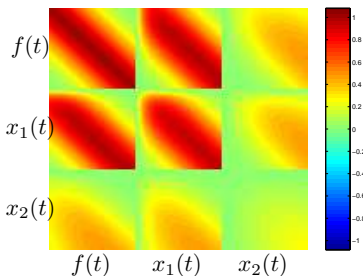
RBF covariance function for $f(t)$

$$x_i(t) = \frac{B_i}{D_i} + S_i \exp(-D_i t) \int_0^t f(u) \exp(D_i u) du.$$

- Joint distribution for $x_1(t)$, $x_2(t)$ and $f(t)$.

► Here:

D_1	S_1	D_2	S_2
5	5	0.5	0.5



► Skip SIM Samples

Joint Sampling of $x(t)$ and $f(t)$ from Covariance

gpsimTest

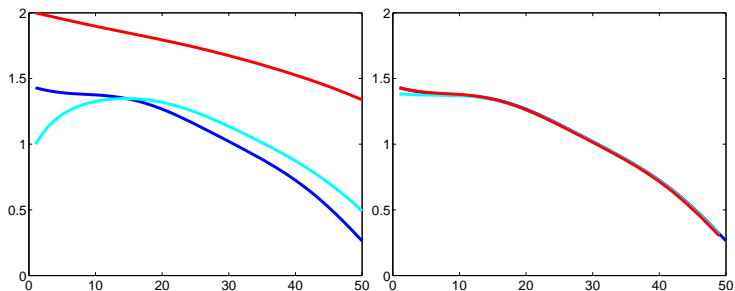


Figure: *Left:* joint samples from the transcription covariance, *blue:* $f(t)$, *cyan:* $x_1(t)$ and *red:* $x_2(t)$. *Right:* numerical solution for $f(t)$ of the differential equation from $x_1(t)$ and $x_2(t)$ (blue and cyan). True $f(t)$ included for comparison.

Joint Sampling of $x(t)$ and $f(t)$ from Covariance

gpsimTest

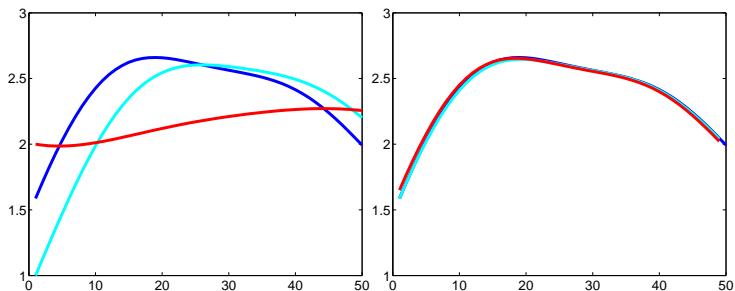


Figure: *Left:* joint samples from the transcription covariance, *blue:* $f(t)$, *cyan:* $x_1(t)$ and *red:* $x_2(t)$. *Right:* numerical solution for $f(t)$ of the differential equation from $x_1(t)$ and $x_2(t)$ (blue and cyan). True $f(t)$ included for comparison.

Joint Sampling of $x(t)$ and $f(t)$ from Covariance

gpsimTest

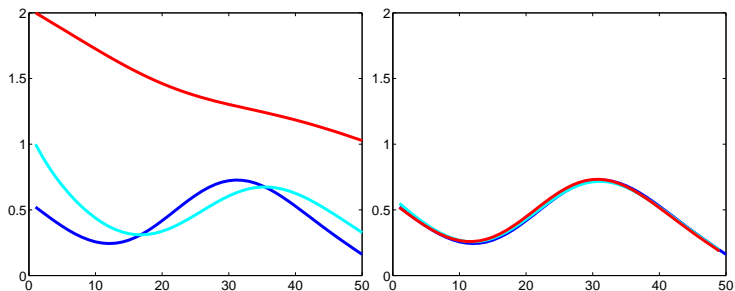


Figure: *Left:* joint samples from the transcription covariance, *blue:* $f(t)$, *cyan:* $x_1(t)$ and *red:* $x_2(t)$. *Right:* numerical solution for $f(t)$ of the differential equation from $x_1(t)$ and $x_2(t)$ (blue and cyan). True $f(t)$ included for comparison.

Any linear operation of a GP \implies Related GP

$$f(t) \sim \mathcal{GP}(0, k_{ff}(t, t')) \implies x_j(t) \sim \mathcal{GP}\left(\frac{B_j}{D_j}, k_{xx}(t, t')\right)$$

Hence, the cross-covariances between the genes is

$$k_{x_i, x_j}(t, t') = S_i S_j \int_0^t \int_0^{t'} e^{-D_i(t-u) - D_j(t'-u')} k_{f,f}(t, t') du du' .$$

Cross-covariances between $x_j(t)$ and $f(t)$ is

$$k_{x_j, f}(t, t') = \int_0^t e^{-D_i(t-u)} k_{f,f}(t, t') du .$$

Prediction of the transcription factor concentration $f(t)$

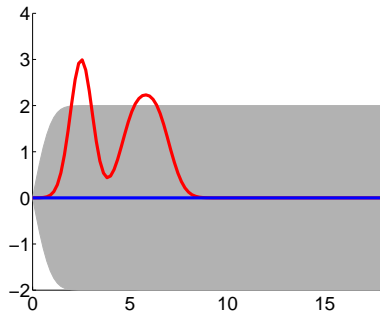
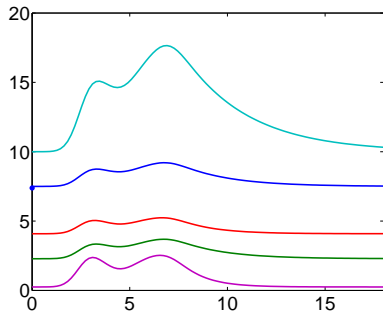
Under the linear model, we have

$$\begin{bmatrix} f \\ \mathbf{x} \end{bmatrix} \sim \mathcal{N} \left(\begin{bmatrix} 0 \\ \frac{\mathbf{B}}{\mathbf{D}} \end{bmatrix}, \begin{bmatrix} K_{ff} & K_{f\mathbf{x}} \\ K_{\mathbf{x}f} & K_{\mathbf{xx}} \end{bmatrix} \right)$$

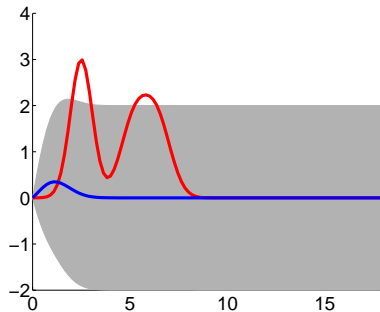
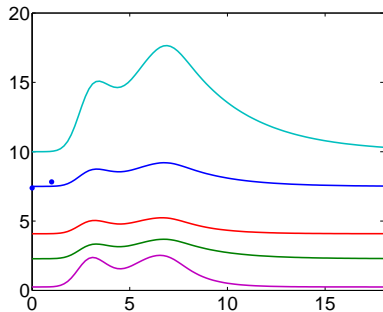
Standard GP Regression yields the mean and covariance function of the predicted process as

$$\begin{aligned} \langle f \rangle_{post} &= K_{f\mathbf{x}} K_{\mathbf{xx}}^{-1} \left(\mathbf{x} - \frac{\mathbf{B}}{\mathbf{D}} \right) \\ K_{ff}^{post} &= K_{ff} - K_{f\mathbf{x}} K_{\mathbf{xx}}^{-1} K_{\mathbf{x}f} \end{aligned}$$

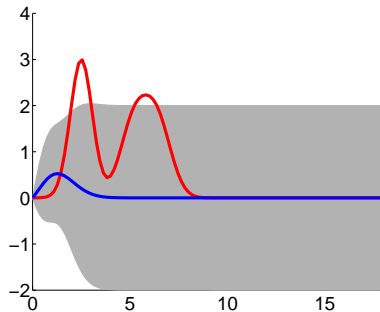
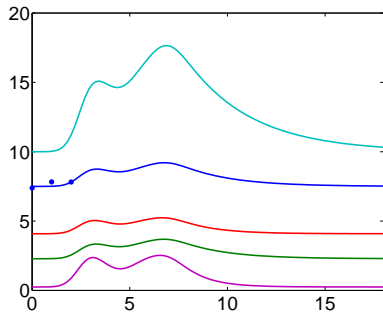
Artificial Example: Inferring $f(t)$



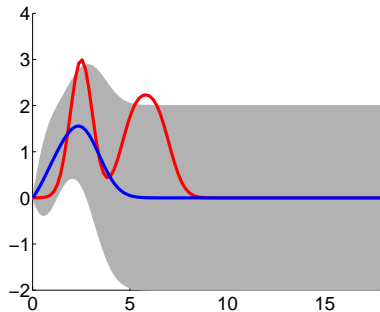
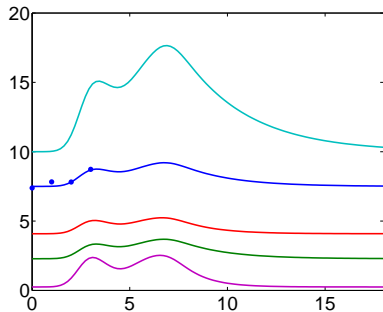
Artificial Example: Inferring $f(t)$



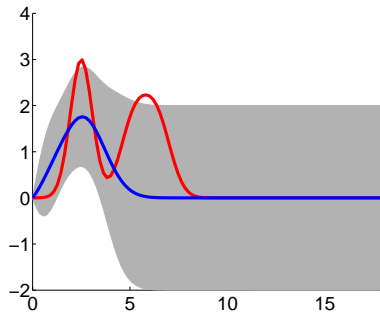
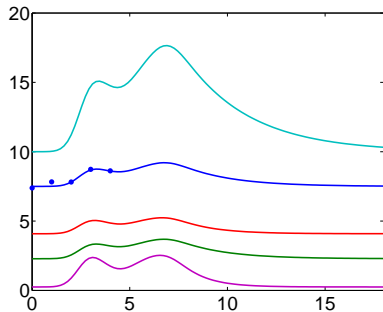
Artificial Example: Inferring $f(t)$



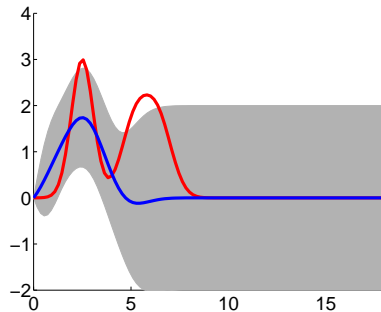
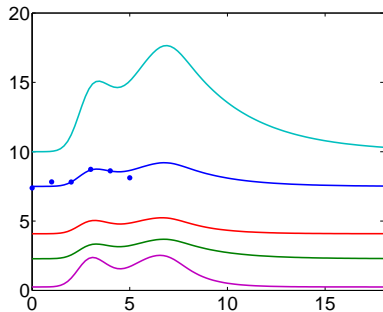
Artificial Example: Inferring $f(t)$



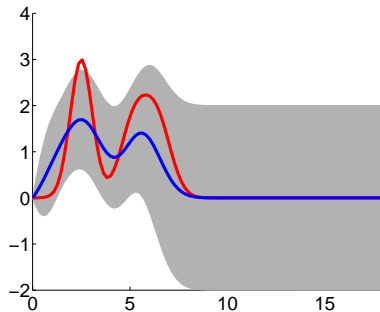
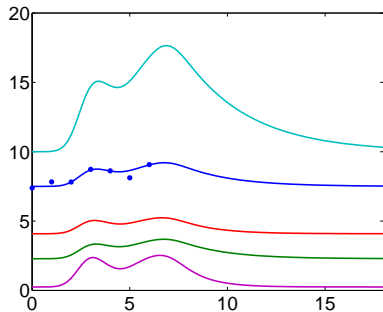
Artificial Example: Inferring $f(t)$



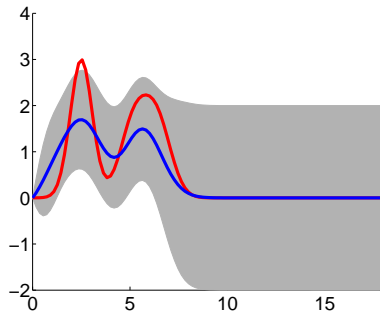
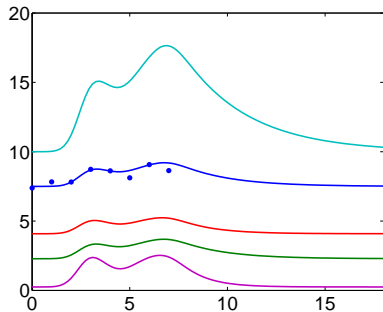
Artificial Example: Inferring $f(t)$



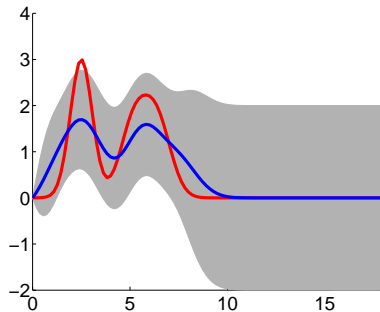
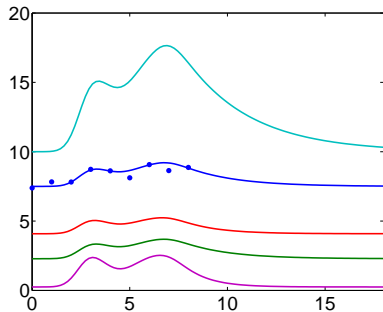
Artificial Example: Inferring $f(t)$



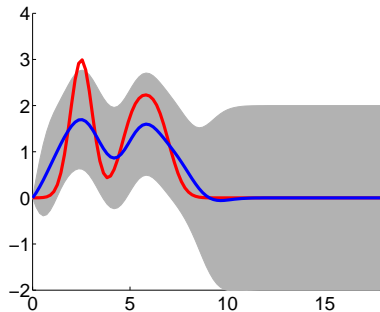
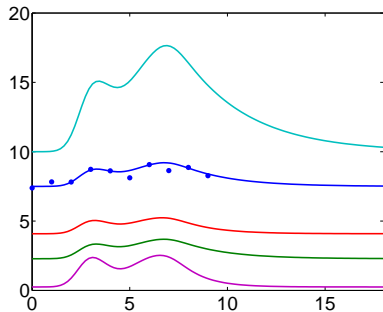
Artificial Example: Inferring $f(t)$



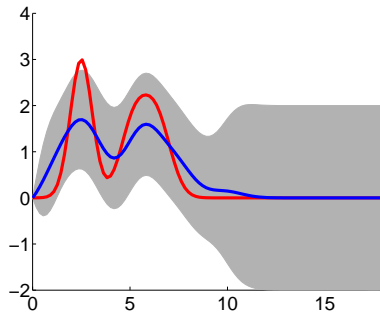
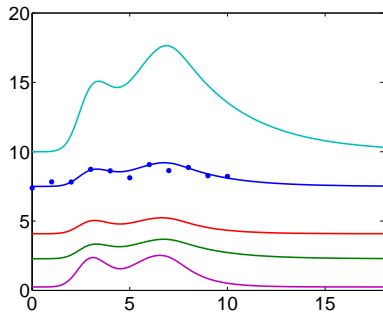
Artificial Example: Inferring $f(t)$



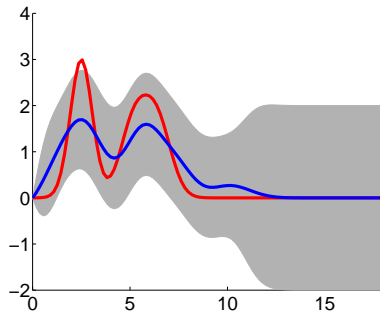
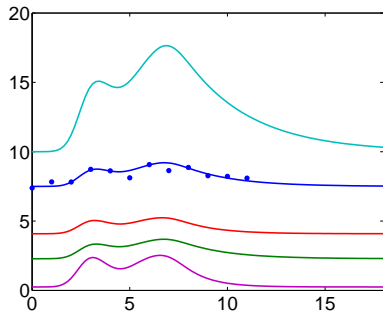
Artificial Example: Inferring $f(t)$



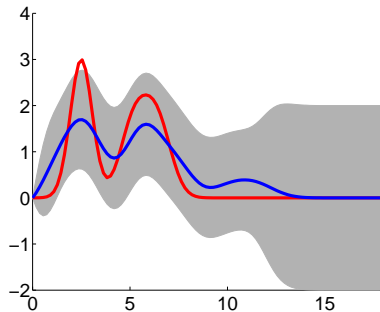
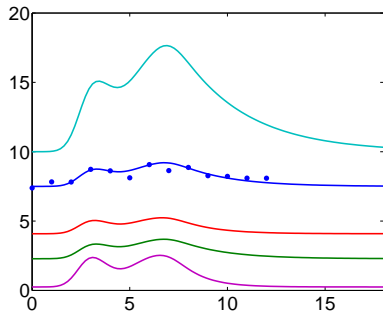
Artificial Example: Inferring $f(t)$



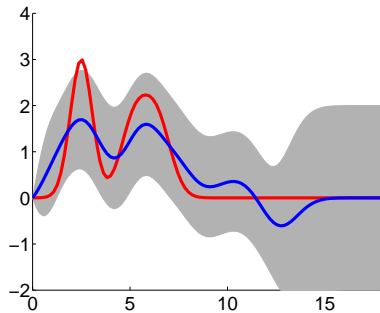
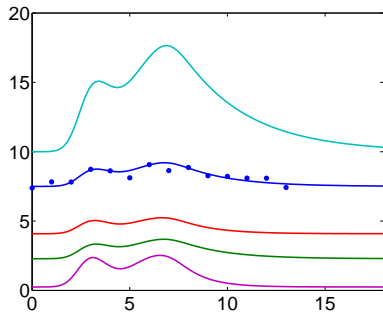
Artificial Example: Inferring $f(t)$



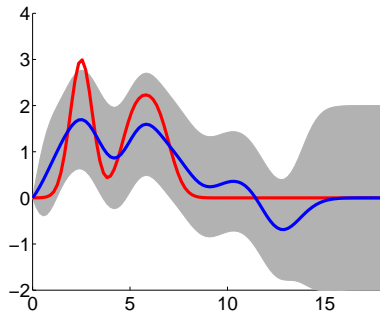
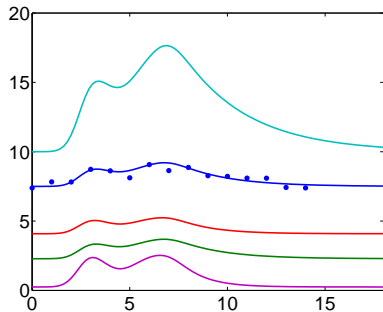
Artificial Example: Inferring $f(t)$



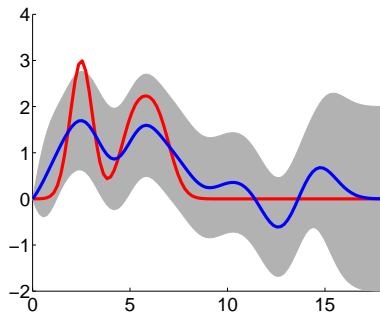
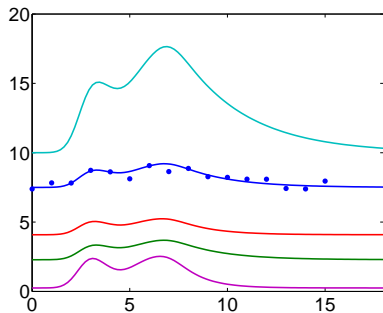
Artificial Example: Inferring $f(t)$



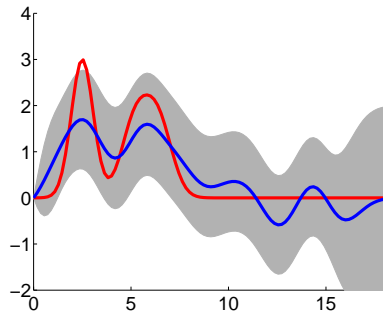
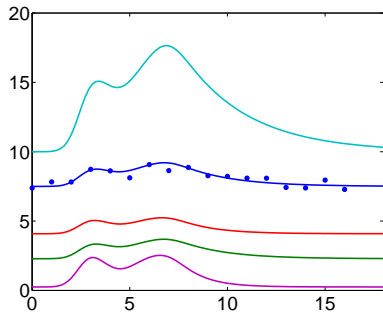
Artificial Example: Inferring $f(t)$



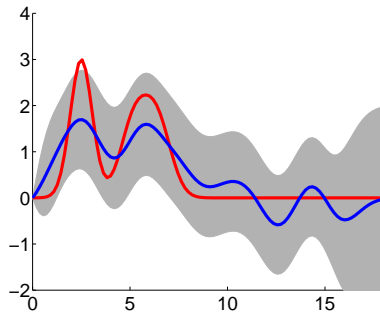
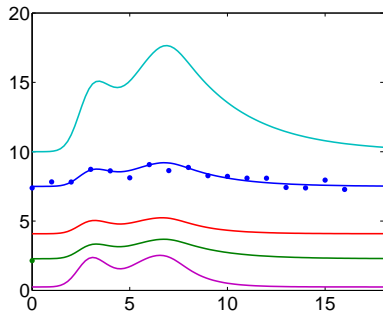
Artificial Example: Inferring $f(t)$



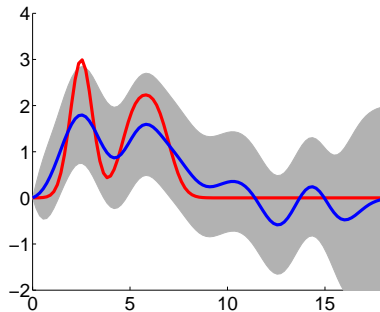
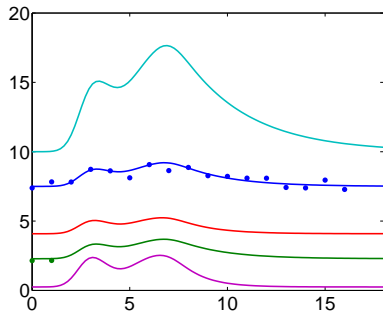
Artificial Example: Inferring $f(t)$



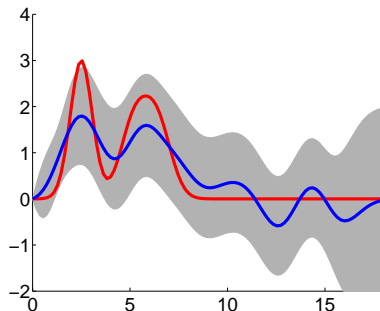
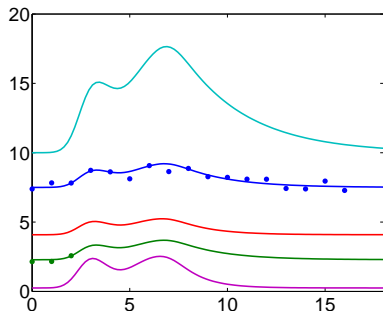
Artificial Example: Inferring $f(t)$



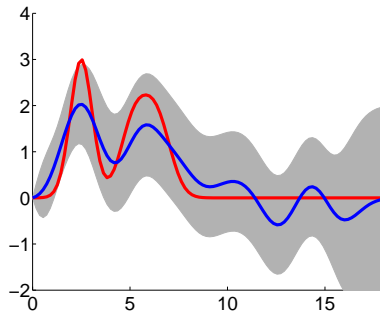
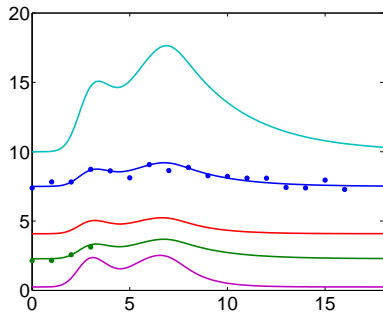
Artificial Example: Inferring $f(t)$



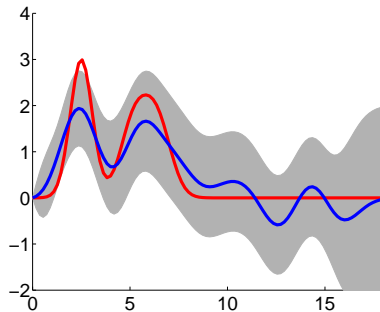
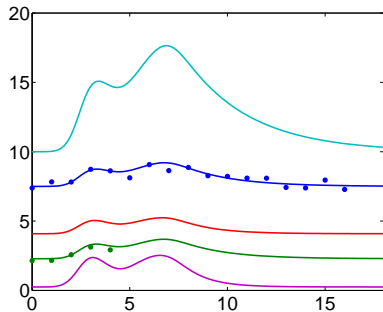
Artificial Example: Inferring $f(t)$



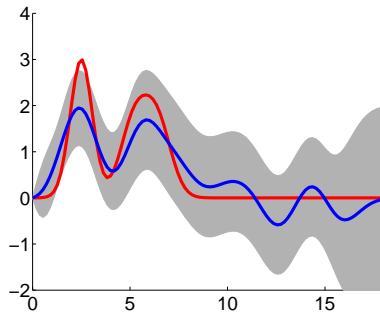
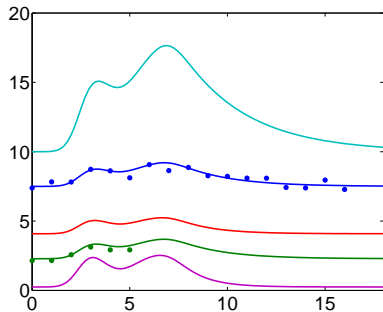
Artificial Example: Inferring $f(t)$



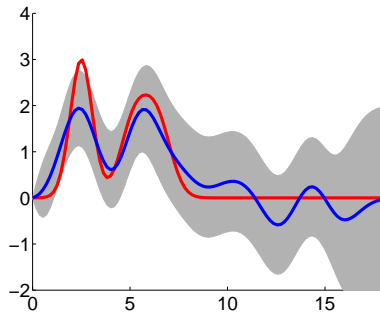
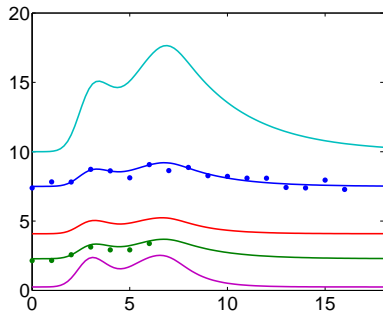
Artificial Example: Inferring $f(t)$



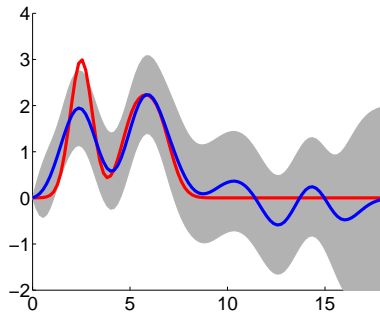
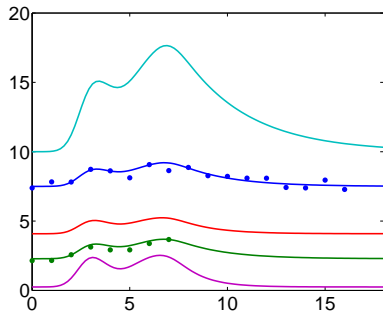
Artificial Example: Inferring $f(t)$



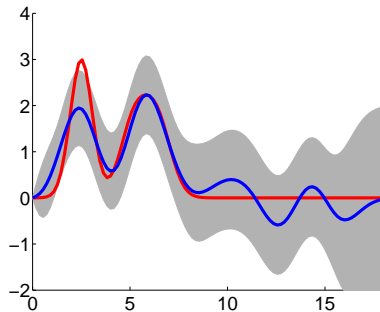
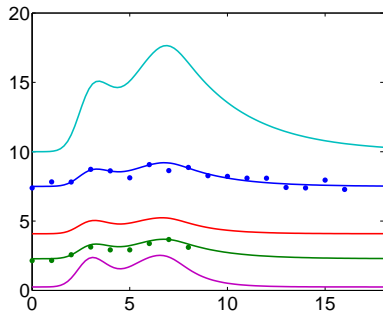
Artificial Example: Inferring $f(t)$



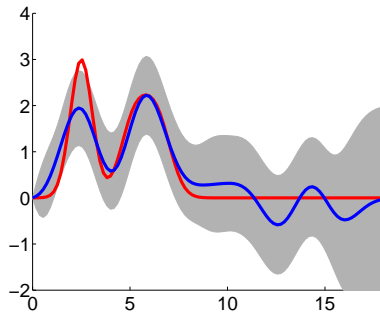
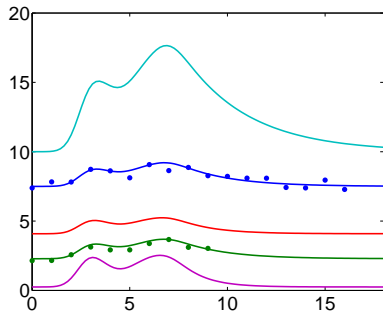
Artificial Example: Inferring $f(t)$



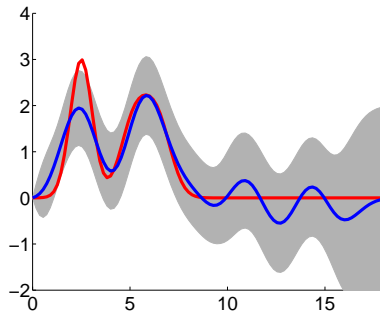
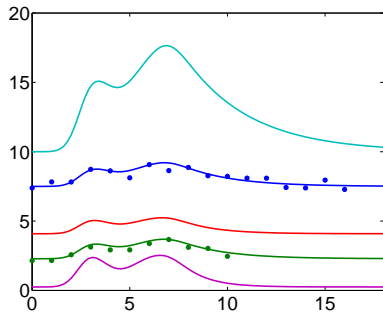
Artificial Example: Inferring $f(t)$



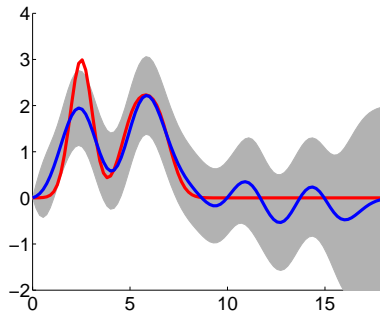
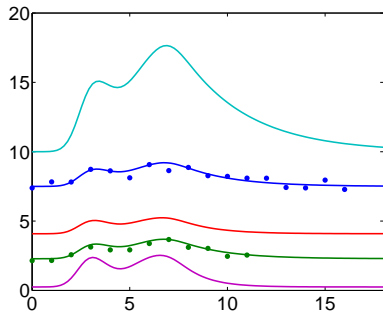
Artificial Example: Inferring $f(t)$



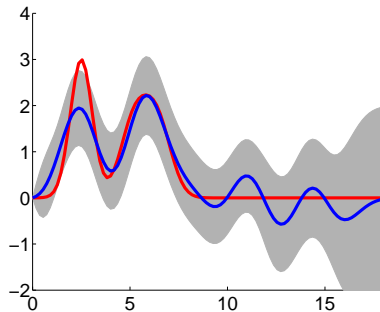
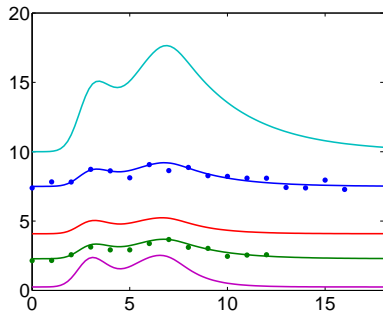
Artificial Example: Inferring $f(t)$



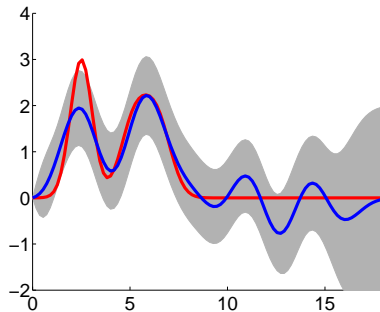
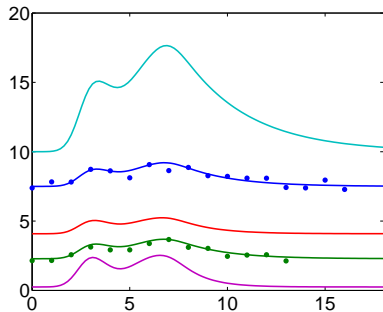
Artificial Example: Inferring $f(t)$



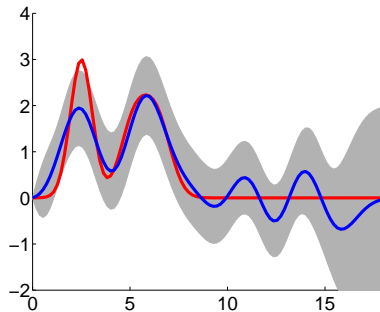
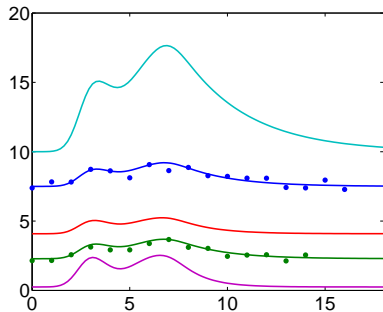
Artificial Example: Inferring $f(t)$



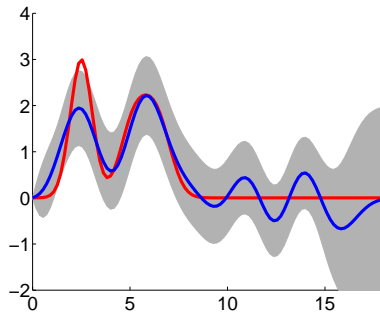
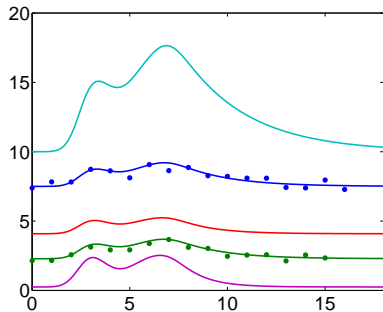
Artificial Example: Inferring $f(t)$



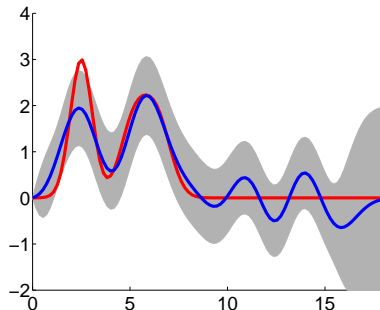
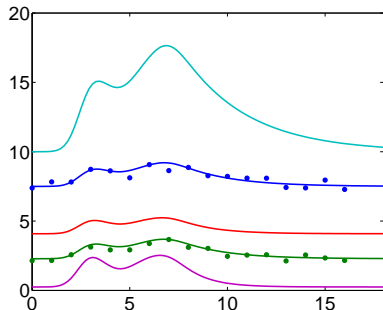
Artificial Example: Inferring $f(t)$



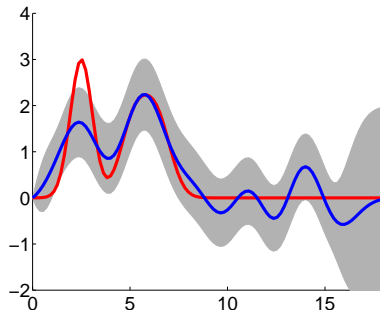
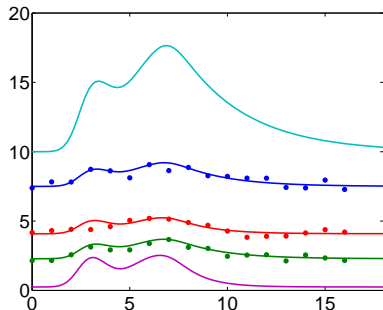
Artificial Example: Inferring $f(t)$



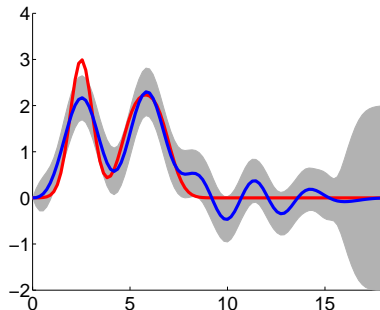
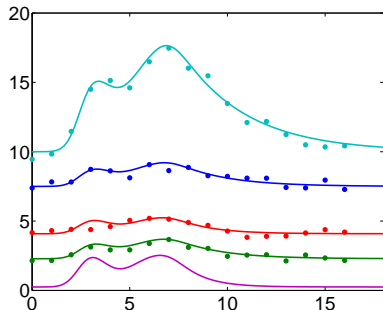
Artificial Example: Inferring $f(t)$



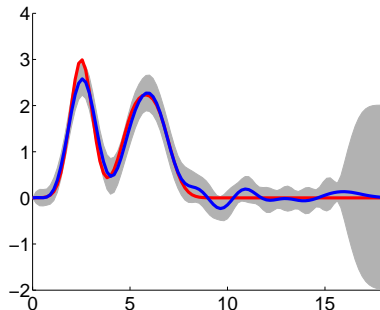
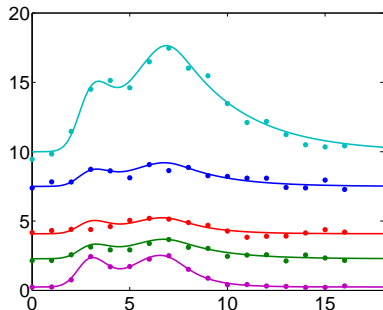
Artificial Example: Inferring $f(t)$



Artificial Example: Inferring $f(t)$



Artificial Example: Inferring $f(t)$



Parameter Estimation for the Linear Model

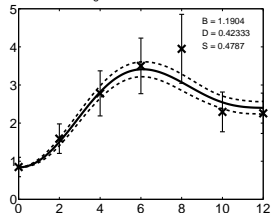
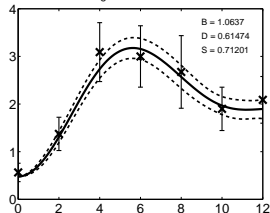
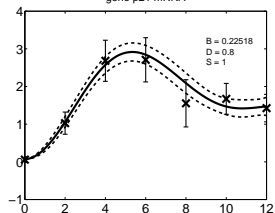
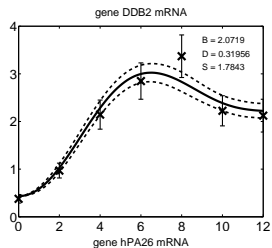
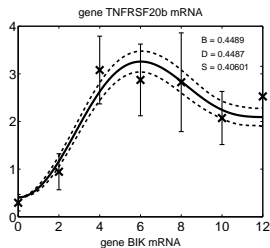
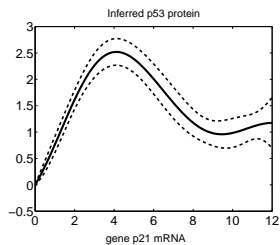
A likelihood function for the model parameters $\theta = \{B_j, S_j, D_j\}_{j=1}^N$ and GP length scale l is obtained by *integrating out* the latent function $f(t)$

$$L(\theta, l) = \int \left(\prod_j p(x_j | \theta, f(t)) \right) p(f(t) | l) df(t)$$

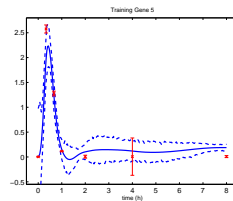
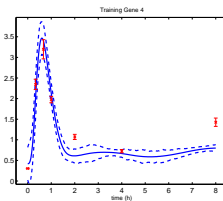
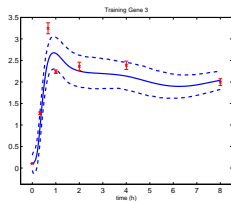
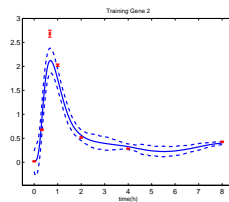
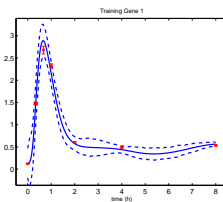
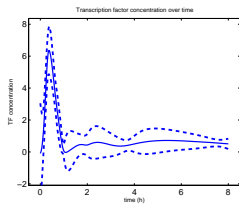
Under the GP model, the log marginal likelihood is then given by

$$\log L(\theta, l) = -\frac{1}{2} x^T (K + \sigma_n^2 \mathbf{I})^{-1} x - \frac{1}{2} \log |K + \sigma_n^2 \mathbf{I}| - \frac{n}{2} \log 2\pi$$

Maximise to find model parameters.

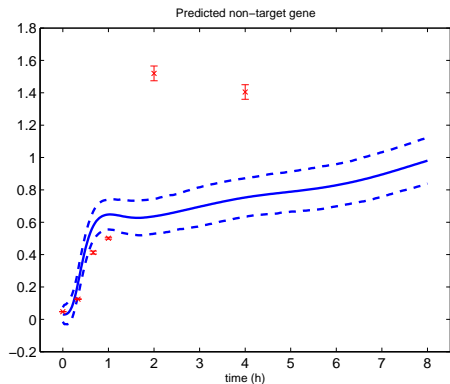
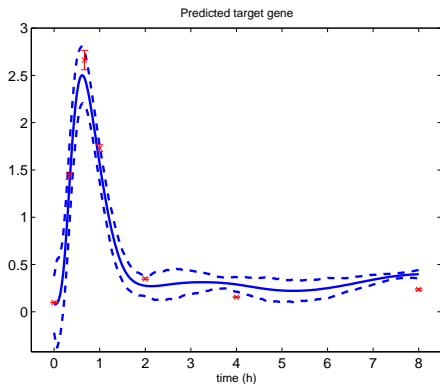


- Target Ranking for Elk-1.
- Elk-1 is phosphorylated by ERK from the EGF signalling pathway.
- Predict concentration of Elk-1 from known targets.
- Rank other targets of Elk-1.



Elk-1 target selection

Fitted model used to rank potential targets of Elk-1



Roadmap

- 1 GPs and Differential Equations
- 2 Cascaded Differential Equations
- 3 Non-linear Response Models
- 4 Discussion and Future Work
- 5 Acknowledgements

Antti Honkela

- Transcription factor protein also has governing mRNA.
- This mRNA can be measured.
- In signalling systems this measurement can be misleading because it is activated (phosphorylated) transcription factor that counts.
- In development phosphorylation plays less of a role.

Collaboration with Furlong Lab in EMBL Heidelberg.

- Mesoderm development in *Drosophila melanogaster* (fruit fly).
- Mesoderm forms in triploblastic animals (along with ectoderm and endoderm). Mesoderm develops into muscles, and circulatory system.
- The transcription factor Twist initiates *Drosophila* mesoderm development, resulting in the formation of heart, somatic muscle, and other cell types.
- Wildtype microarray experiments publicly available.
- Can we use the cascade model to predict viable targets of Twist?

We take the production rate of active transcription factor to be given by

$$\begin{aligned}\frac{df(t)}{dt} &= \sigma y(t) - \delta f(t) \\ \frac{dx_j(t)}{dt} &= B_j + S_j f(t) - D_j x_j(t)\end{aligned}$$

The solution for $f(t)$, setting transient terms to zero, is

$$f(t) = \sigma \exp(-\delta t) \int_0^t y(u) \exp(\delta u) du .$$

Covariance for Translation/Transcription Model

RBF covariance function for $y(t)$

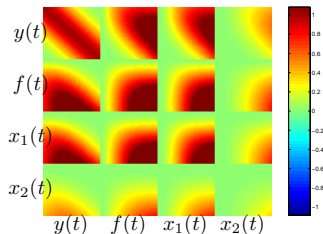
$$f(t) = \sigma \exp(-\delta t) \int_0^t y(u) \exp(\delta u) du$$

$$x_i(t) = \frac{B_i}{D_i} + S_i \exp(-D_i t) \int_0^t f(u) \exp(D_i u) du.$$

- Joint distribution for $x_1(t)$, $x_2(t)$, $f(t)$ and $y(t)$.

- Here:

δ	D_1	S_1	D_2	S_2
0.1	5	5	0.5	0.5



- Use mRNA of Twist as driving input.
- For each gene build a cascade model that forces Twist to be the only TF.
- Compare fit of this model to a baseline (e.g. similar model but sensitivity zero).
- Rank according to the likelihood above the baseline.

Results for Twi using the Cascade model

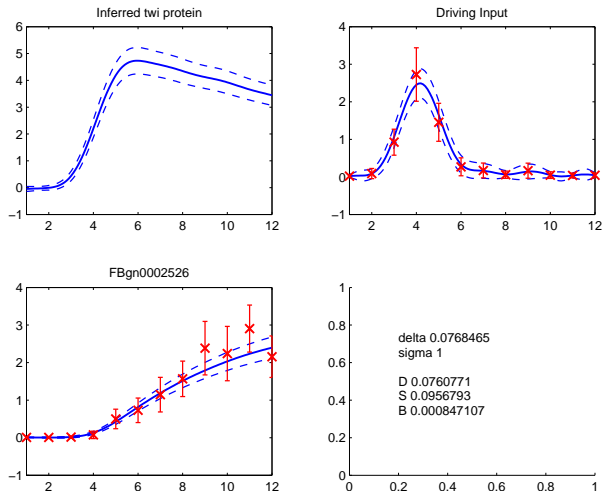


Figure: Model for flybase gene identity FBgn0002526.

Results for Twi using the Cascade model

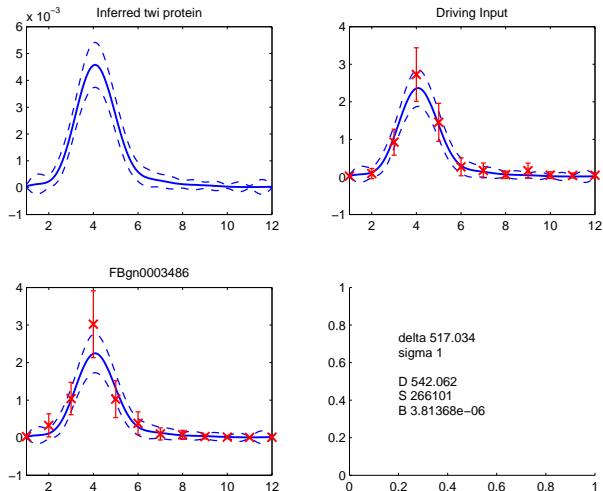


Figure: Model for flybase gene identity FBgn0003486.

Results for Twi using the Cascade model

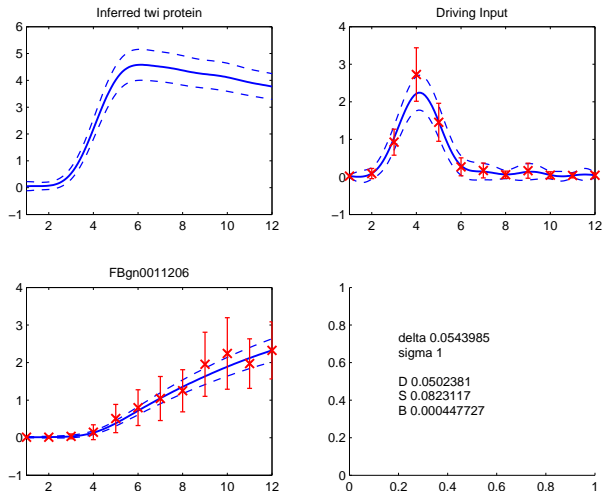


Figure: Model for flybase gene identity FBgn0011206.

Results for Twi using the Cascade model

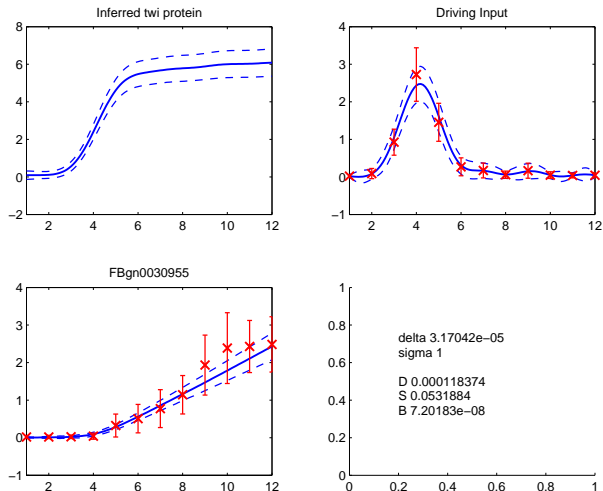


Figure: Model for flybase gene identity FBgn0030955.

Results for Twi using the Cascade model

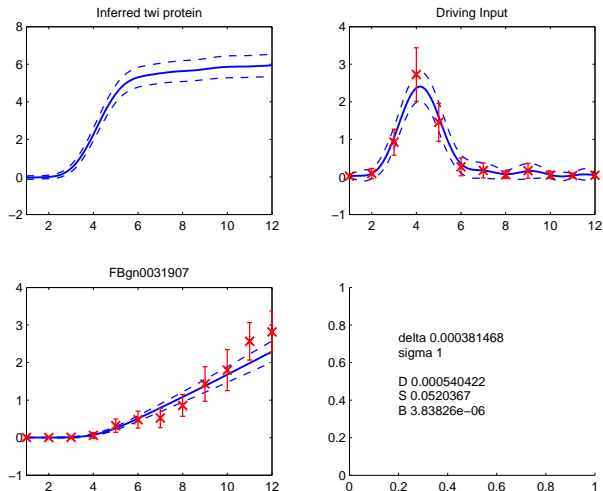


Figure: Model for flybase gene identity FBgn0031907.

Results for Twi using the Cascade model

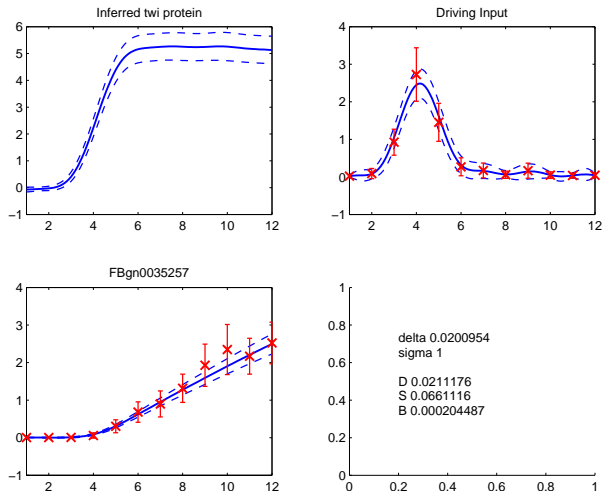


Figure: Model for flybase gene identity FBgn0035257.

Results for Twi using the Cascade model

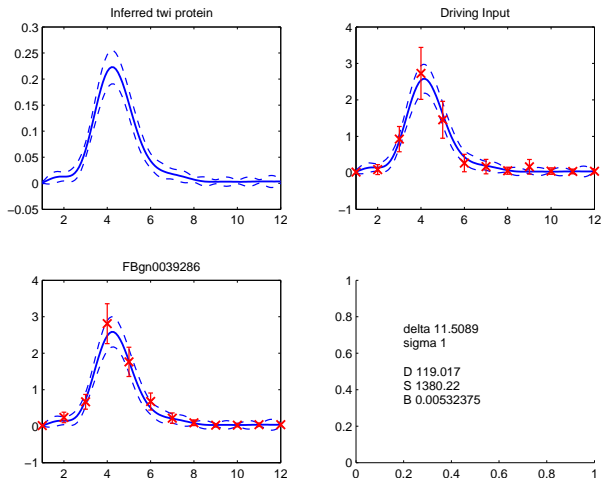


Figure: Model for flybase gene identity FBgn0039286.

Results of Ranking

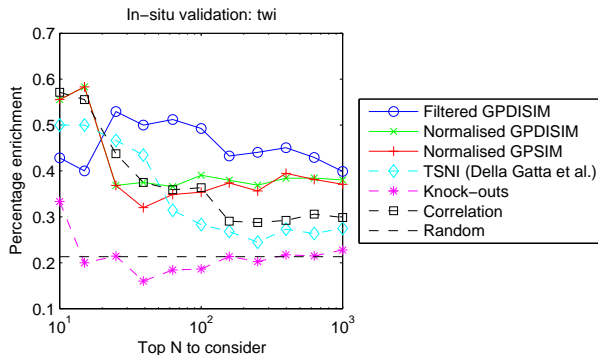


Figure: Percentage enrichment for top N targets for relevant terms in *Drosophila* in situ.

Results of Ranking

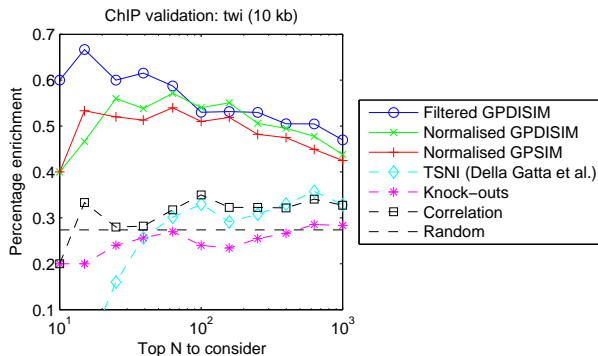


Figure: Percentage enrichment for top N targets for ChIP-chip confirmed targets.

- Cascade models allow genomewide analysis of potential targets given only expression data.
- Once a set of potential candidate targets have been identified, they can be modelled in a more complex manner.
- We don't have ground truth, but evidence indicates that the approach *can* perform as well as knockouts.

Roadmap

- 1 GPs and Differential Equations
- 2 Cascaded Differential Equations
- 3 Non-linear Response Models**
- 4 Discussion and Future Work
- 5 Acknowledgements

Consider the following modification to the model,

$$\frac{dx_j(t)}{dt} = B_j + S_j g(f(t)) - D_j x_j(t),$$

where $g(\cdot)$ is a non-linear function. The differential equation can still be solved,

$$x_j(t) = \frac{B_j}{D_j} + S_j \int_0^t e^{-D_j(t-u)} g_j(f(u)) du$$

Based on Laplace's method,

$$p(\mathbf{f} \mid \mathbf{x}) = N(\hat{\mathbf{f}}, \mathbf{A}^{-1}) \propto \exp\left(-\frac{1}{2}(\mathbf{f} - \hat{\mathbf{f}})^T \mathbf{A}(\mathbf{f} - \hat{\mathbf{f}})\right)$$

where $\hat{\mathbf{f}} = \operatorname{argmax} p(\mathbf{f} \mid \mathbf{x})$ and $\mathbf{A} = -\nabla \nabla \log p(\mathbf{f} \mid \mathbf{y})|_{\mathbf{f}=\hat{\mathbf{f}}}$ is the Hessian of the negative posterior at that point. To obtain $\hat{\mathbf{f}}$ and \mathbf{A} , we define the

following function $\psi(\mathbf{f})$ as:

$$\log p(\mathbf{f} \mid \mathbf{x}) \propto \psi(\mathbf{f}) = \log p(\mathbf{x} \mid \mathbf{f}) + \log p(\mathbf{f})$$

Assigning a GP prior distribution to $f(t)$, it then follows that

$$\log p(\mathbf{f}) = -\frac{1}{2}\mathbf{f}^T \mathbf{K}^{-1}\mathbf{f} - \frac{1}{2}\log |\mathbf{K}| - \frac{n}{2}\log 2\pi$$

where \mathbf{K} is the covariance matrix of $f(t)$. Hence,

$$\begin{aligned}\nabla\psi(\mathbf{f}) &= \nabla \log p(\mathbf{x}|\mathbf{f}) - \mathbf{K}^{-1}\mathbf{f} \\ \nabla\nabla\psi(\mathbf{f}) &= \nabla\nabla \log p(\mathbf{x}|\mathbf{f}) - \mathbf{K}^{-1} = -\mathbf{W} - \mathbf{K}^{-1}\end{aligned}$$

Estimation of $\psi(\mathbf{f})$

Newton's method is applied to find the maximum of $\psi(\mathbf{f})$ as

$$\begin{aligned}\mathbf{f}^{new} &= \mathbf{f} - (\nabla \nabla \psi(\mathbf{f}))^{-1} \nabla \psi(\mathbf{f}) \\ &= (\mathbf{W} + \mathbf{K}^{-1})^{-1} (\mathbf{W}\mathbf{f} - \nabla \log p(\mathbf{x}|\mathbf{f}))\end{aligned}$$

In addition, $\mathbf{A} = -\nabla \nabla \psi(\hat{\mathbf{f}}) = \mathbf{W} + \mathbf{K}^{-1}$ where \mathbf{W} is the negative Hessian matrix. Hence, the Laplace approximation to the posterior is a Gaussian with mean $\hat{\mathbf{f}}$ and covariance matrix \mathbf{A}^{-1} as

$$p(\mathbf{f} \mid \mathbf{x}) \simeq N(\hat{\mathbf{f}}, \mathbf{A}^{-1}) = N(\hat{\mathbf{f}}, (\mathbf{W} + \mathbf{K}^{-1})^{-1})$$

Model Parameter Estimation

The marginal likelihood is useful for estimating the model parameters θ and covariance parameters I

$$p(\mathbf{x}|\theta, \phi) = \int p(\mathbf{x}|\mathbf{f}, \theta) p(\mathbf{f}|\phi) d\mathbf{f} = \int \exp(\psi(\mathbf{f})) d\mathbf{f}$$

Using Taylor expansion of $\psi(\mathbf{f})$,

$$\log p(\mathbf{x}|\theta, \phi) = \log p(\mathbf{x}|\hat{\mathbf{f}}, \theta, \phi) - \frac{1}{2}\mathbf{f}^T \mathbf{K}^{-1} \mathbf{f} - \frac{1}{2} \log |\mathbf{I} + \mathbf{K} \mathbf{W}|$$

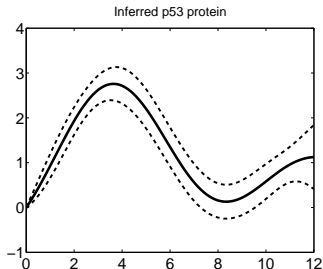
The parameters $\eta = \{\theta, \phi\}$ can be then estimated by using

$$\frac{\partial \log p(\mathbf{x}|\eta)}{\partial \eta} = \frac{\partial \log p(\mathbf{x}|\eta)}{\partial \eta} \Big|_{\text{explicit}} + \frac{\partial \log p(\mathbf{x}|\eta)}{\partial \hat{\mathbf{f}}} \frac{\partial \hat{\mathbf{f}}}{\partial \eta}$$

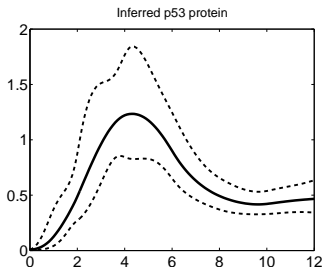
- The Michaelis-Menten activation model uses the following non-linearity

$$g_j(f(t)) = \frac{e^{f(t)}}{\gamma_j + e^{f(t)}},$$

where we are using a GP $f(t)$ to model the log of the TF activity.



(a)



(b)

Validation of Laplace Approximation

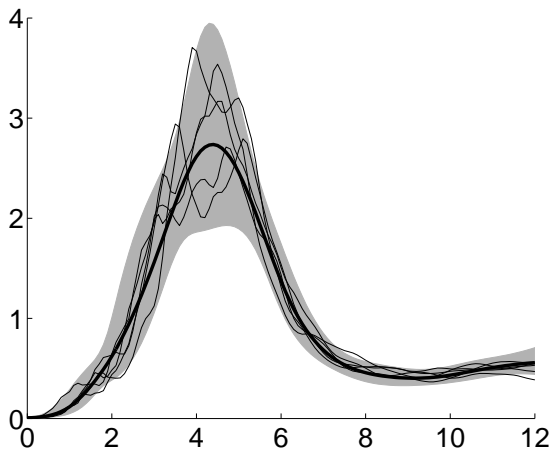


Figure: Laplace approximation error bars along with samples from the true posterior distribution.

- DNA damage may occur as a result of activity of antibiotics.
- LexA is bound to the genome preventing transcription of the SOS genes.
- RecA protein is stimulated by single stranded DNA, inactivates the LexA repressor.
- This allows several of the LexA targets to transcribe.
- The SOS pathway may be essential in antibiotic resistance Cirz et al. (2005).
- Aim is to target these proteins to produce drugs to increase efficacy of antibiotics Lee et al. (2005).

- Data from Courcelle et al. (2001)
- UV irradiation of *E. coli*. in both wild-type cells and *lexA1* mutants, which are unable to induce genes under LexA control.
- Response measured with two color hybridization to cDNA arrays.

Their Model

Given measurements of gene expression at N time points $(t_0, t_1, \dots, t_{N-1})$, the temporal profile of a gene i , $x_i(t)$, that solves the ODE in Eq. 1 can be approximated by

$$x_i(t) = x_i^0 e^{-\delta_i t} + \frac{B_i}{D_i} + S_i e^{-\delta_i t} \frac{1}{D_i} \sum_{j=0}^{N-2} (e^{D_i t_{j+1}} - e^{D_i t_j}) \frac{1}{\gamma_i + \bar{f}_j}$$

where $\bar{f}_j = \frac{(f(t_j) + f(t_{j+1}))}{2}$ on each subinterval (t_j, t_{j+1}) , $j = 0, \dots, N-2$. This is under the simplifying assumption that $f(t)$ is a piece-wise constant function on each subinterval (t_j, t_{j+1}) .

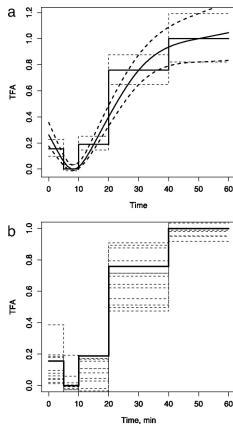


Figure: Fig. 2 from Khanin et al. (2006): Reconstructed activity level of master repressor LexA, following a UV dose of 40 J/m².

Their Results

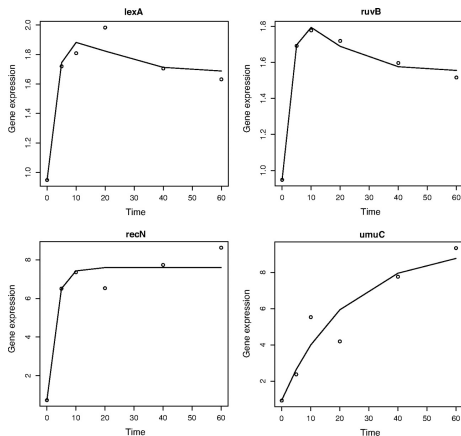


Figure: Fig. 3 from Khanin et al. (2006): Reconstructed profiles for four genes in the LexA SIM.

- We can use the same model of repression,

$$g_j(f(t)) = \frac{1}{\gamma_j + e^{f(t)}}$$

In the case of repression we have to include the transient term,

$$x_j(t) = \alpha_j e^{-D_j t} + \frac{B_j}{D_j} + S_j \int_0^t e^{-D_j(t-u)} g_j(f(u)) du$$

Results for the repressor LexA

Pei Gao

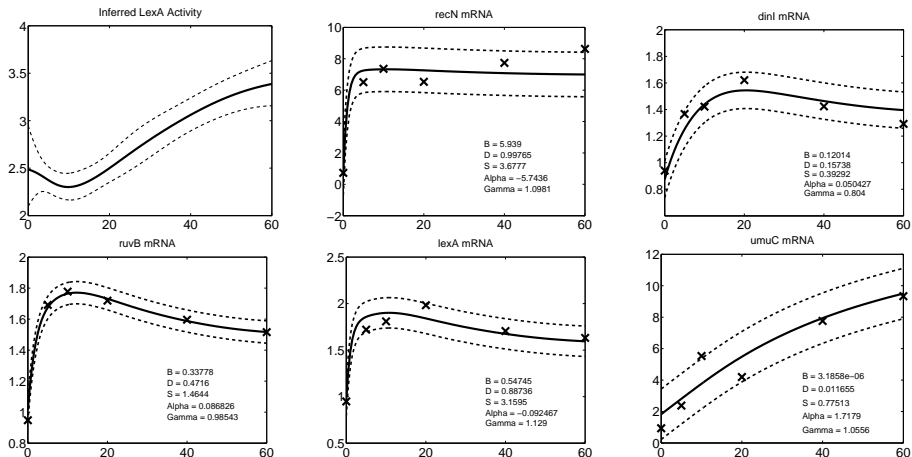


Figure: Our results using an MLP kernel. To appear at ECCB08 Gao et al. (2008).

- Sample in Gaussian processes

$$p(\mathbf{f}|\mathbf{x}) \propto p(\mathbf{x}|\mathbf{f}) p(\mathbf{f})$$

- Likelihood relates GP to data through

$$x_j(t) = \alpha_j e^{-D_j t} + \frac{B_j}{D_j} + S_j \int_0^t e^{-D_j(t-u)} g_j(f(u)) du$$

- We use *control points* for fast sampling.

The Metropolis-Hastings algorithm

- Initialize $\mathbf{f}^{(0)}$
- Form a Markov chain. Use a proposal distribution $Q(\mathbf{f}^{(t+1)}|\mathbf{f}^{(t)})$ and accept with the M-H step

$$\min \left(1, \frac{p(\mathbf{x}|\mathbf{f}^{(t+1)})p(\mathbf{f}^{(t+1)})}{p(\mathbf{x}|\mathbf{f}^{(t)})p(\mathbf{f}^{(t)})} \frac{Q(\mathbf{f}^{(t)}|\mathbf{f}^{(t+1)})}{Q(\mathbf{f}^{(t+1)}|\mathbf{f}^{(t)})} \right)$$

- \mathbf{f} can be very *high dimensional* (hundreds of points)
- How do we choose the proposal $Q(\mathbf{f}^{(t+1)}|\mathbf{f}^{(t)})$?
 - ▶ Can we use the GP prior $p(\mathbf{f})$ as the proposal?

Sampling using control points

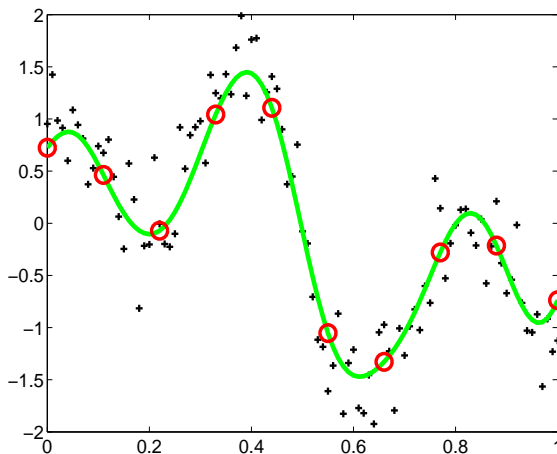
- Separate the points in \mathbf{f} into two groups:
 - ▶ few control points \mathbf{f}_c
 - ▶ and the large majority of the remaining points $\mathbf{f}_\rho = \mathbf{f} \setminus \mathbf{f}_c$
- Sample the control points \mathbf{f}_c using a proposal $q\left(\mathbf{f}_c^{(t+1)}|\mathbf{f}_c^{(t)}\right)$
- Sample the remaining points \mathbf{f}_ρ using the conditional GP prior $p\left(\mathbf{f}_\rho^{(t+1)}|\mathbf{f}_c^{(t+1)}\right)$
- The whole proposal is

$$Q\left(\mathbf{f}^{(t+1)}|\mathbf{f}^{(t)}\right) = p\left(\mathbf{f}_\rho^{(t+1)}|\mathbf{f}_c^{(t+1)}\right) q\left(\mathbf{f}_c^{(t+1)}|\mathbf{f}_c^{(t)}\right)$$

- Its like sampling from the prior $p(\mathbf{f})$ but imposing random walk behaviour through the control points

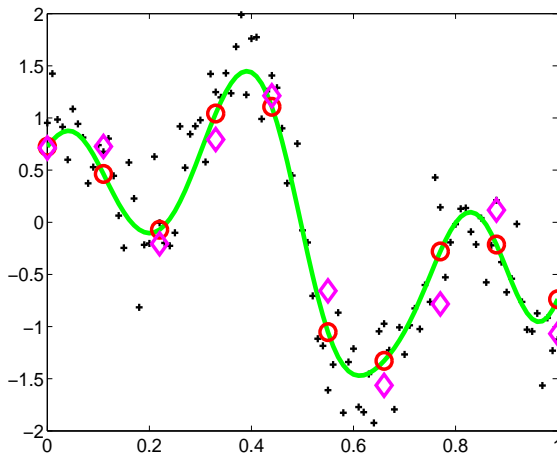
Sampling using control points: Regression-Examples

Sample 121 points using 10 control points



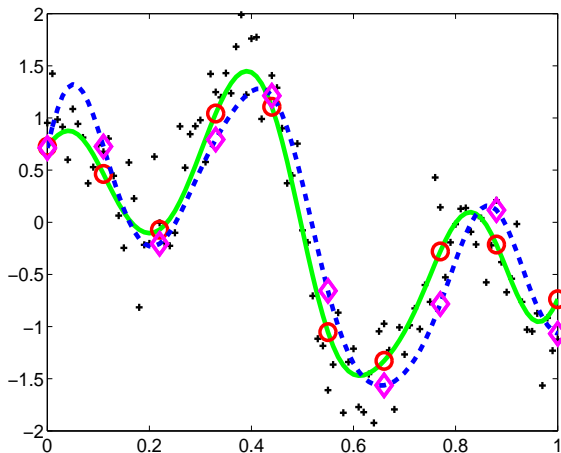
Sampling using control points: Regression-Examples

Sample 121 points using 10 control points



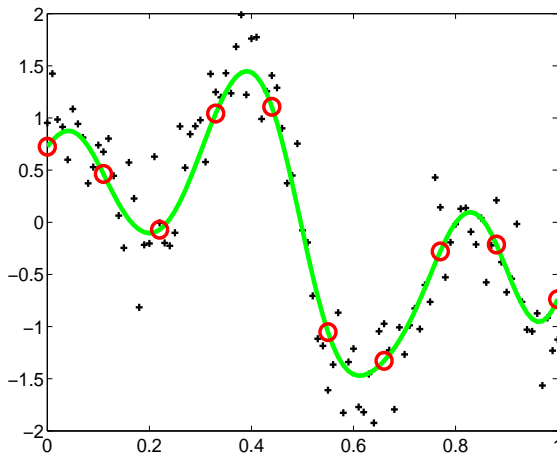
Sampling using control points: Regression-Examples

Sample 121 points using 10 control points



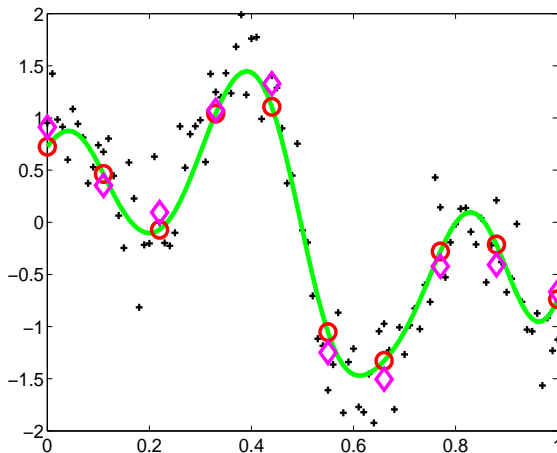
Sampling using control points: Regression-Examples

Sample 121 points using 10 control points



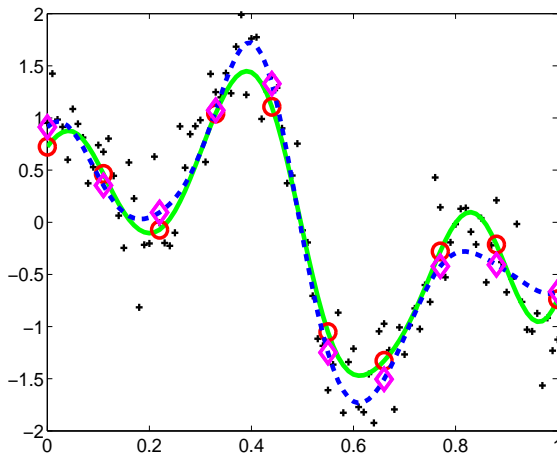
Sampling using control points: Regression-Examples

Sample 121 points using 10 control points



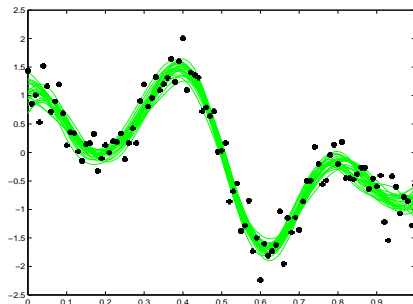
Sampling using control points: Regression-Examples

Sample 121 points using 10 control points



Sampling using control points

Few samples drawn during MCMC

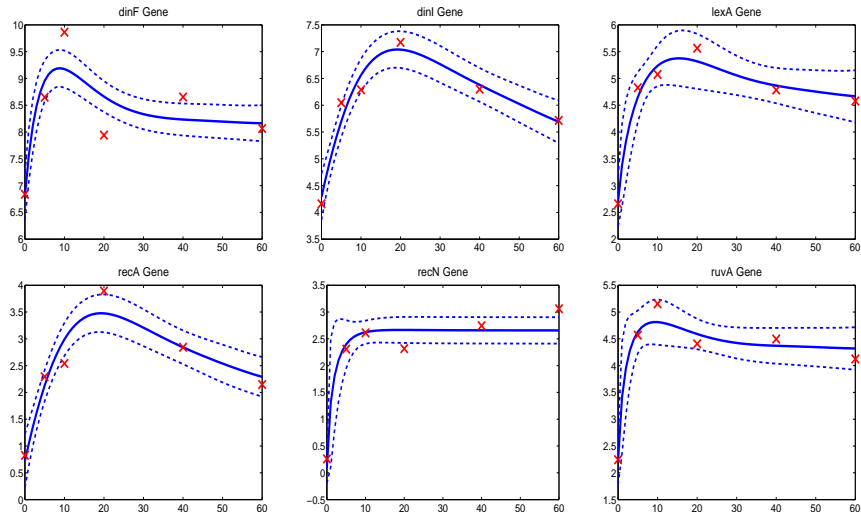


- Again consider the Michaelis-Menten kinetic equation

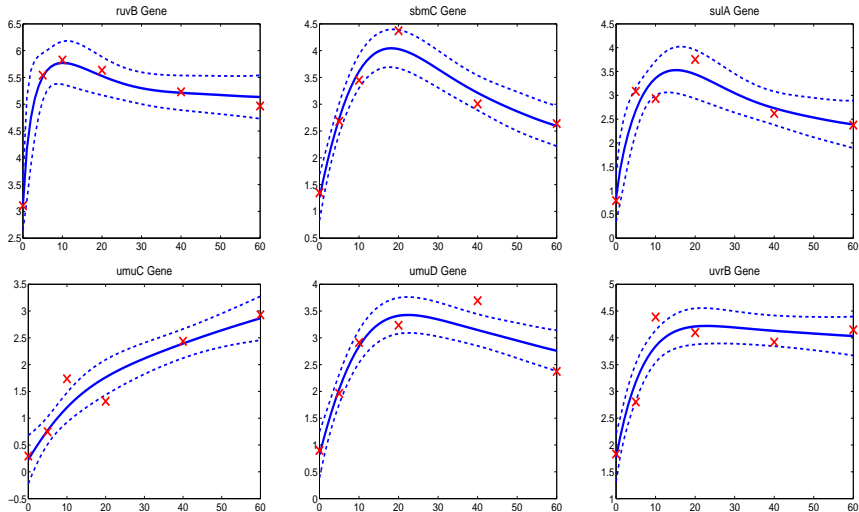
$$\frac{dx_j(t)}{dt} = B_j + S_j \frac{1}{\exp(f(t)) + \gamma_j} - D_j x_j(t)$$

- We have 14 genes (5 kinetic parameters each)
- Gene expressions are available for $T = 6$ time slots
- TF (\mathbf{f}) is discretized using 121 points
- MCMC details:
 - ▶ 6 control points are used (placed in a equally spaced grid)
 - ▶ Running time was 5 hours for 2 million sampling iterations plus burn in
 - ▶ Acceptance rate for \mathbf{f} after burn in was between 15% – 25%

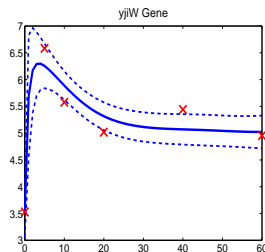
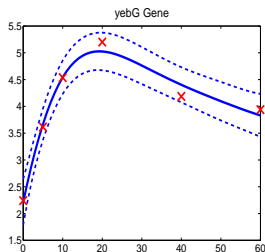
Results in E.coli data: Predicted gene expressions



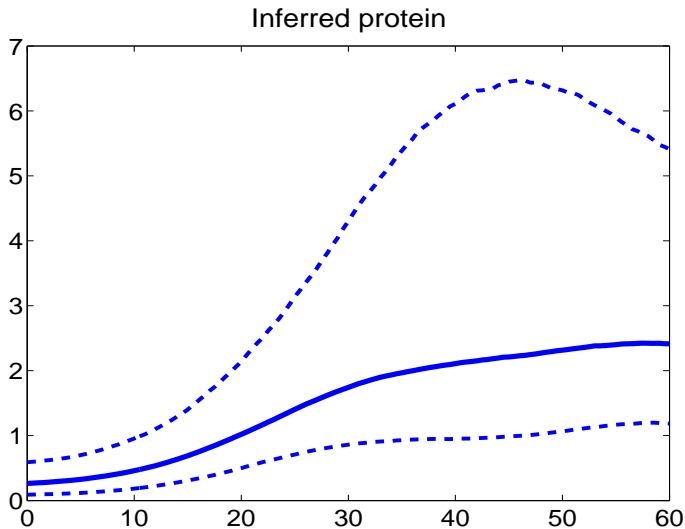
Results in E.coli data: Predicted gene expressions



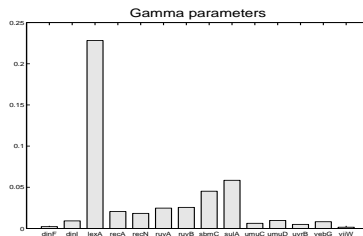
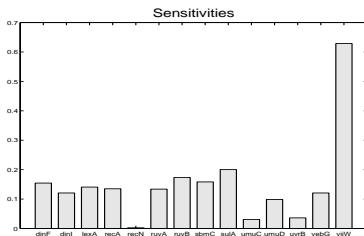
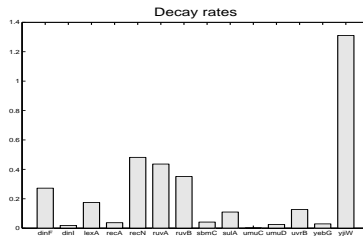
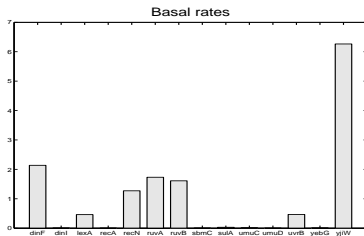
Results in E.coli data: Predicted gene expressions



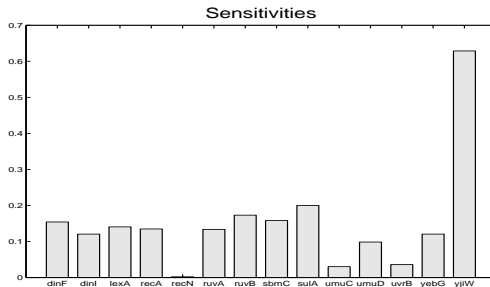
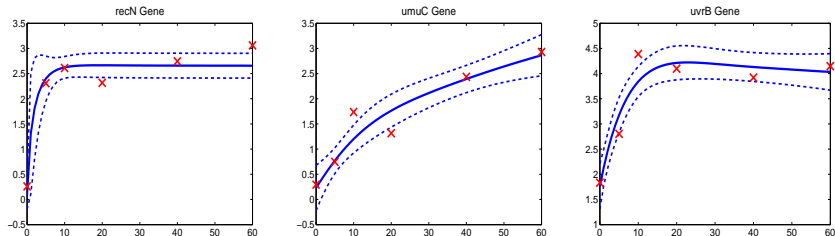
Results in E.coli data: Protein concentration



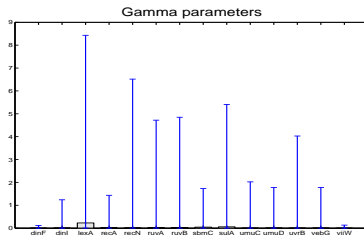
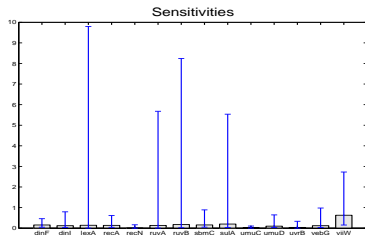
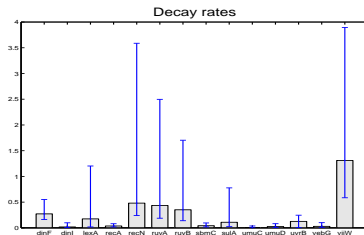
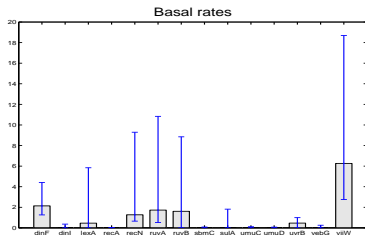
Results in E.coli data: Kinetic parameters



Results in E.coli data: Genes with low sensitivity value



Results in E.coli data: Confidence intervals for the kinetic parameters

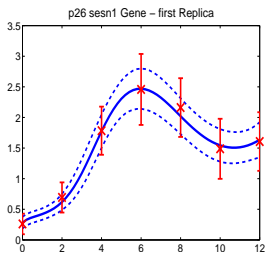
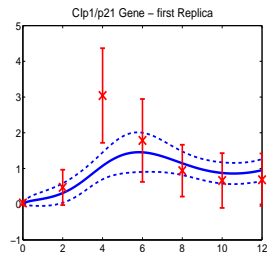
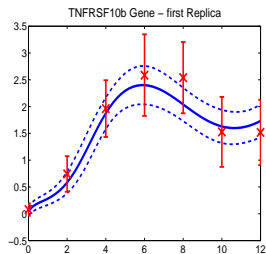
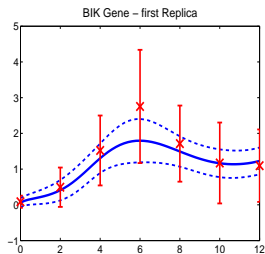
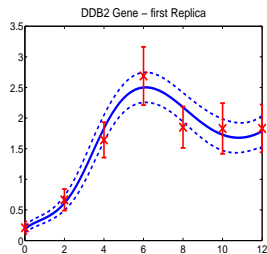


- One transcription factor (p53) that acts as an activator. We consider the Michaelis-Menten kinetic equation

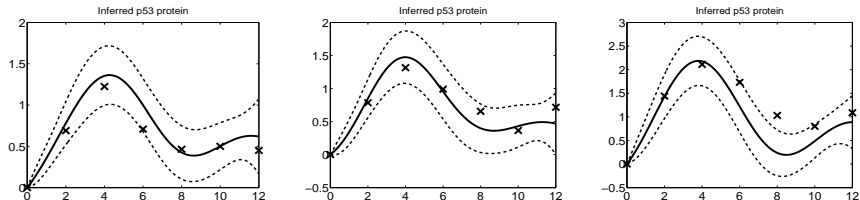
$$\frac{dx_j(t)}{dt} = B_j + S_j \frac{\exp(f(t))}{\exp(f(t)) + \gamma_j} - D_j x_j(t)$$

- We have 5 genes
- Gene expressions are available for $T = 7$ times and there are 3 replicas of the time series data
- TF (\mathbf{f}) is discretized using 121 points
- MCMC details:
 - ▶ 7 control points are used (placed in a equally spaced grid)
 - ▶ Running time 4/5 hours for 2 million sampling iterations plus burn in
 - ▶ Acceptance rate for \mathbf{f} after burn in was between 15% – 25%

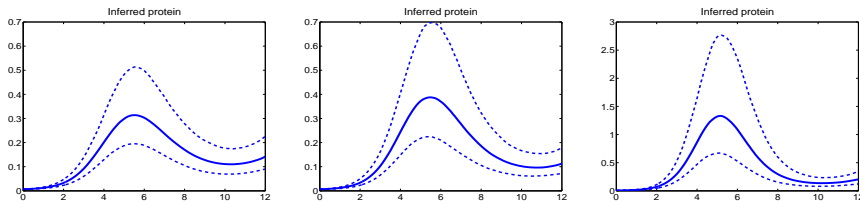
Data used by Barenco et al. (2006): Predicted gene expressions for the 1st replica



Data used by Barenco et al. (2006): Protein concentrations

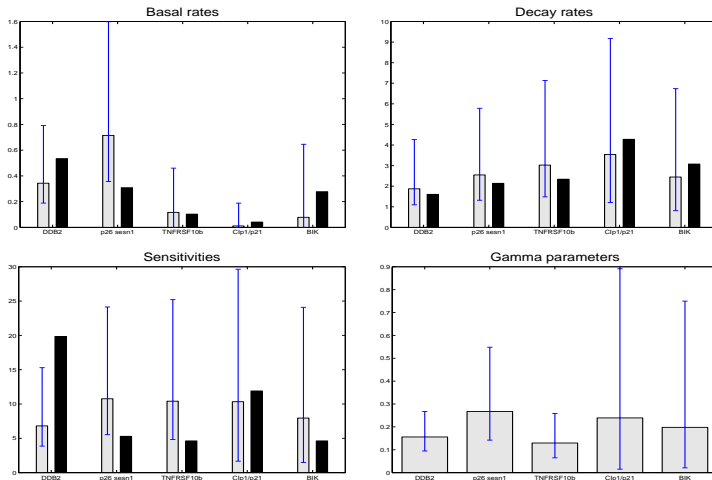


Linear model (Barenco et al. predictions are shown as crosses)



Nonlinear (Michaelis-Menten kinetic equation)

p53 Data Kinetic parameters



Our results (grey) compared with Barenco et al. (2006) (black). Note that Barenco et al. use a linear model

Roadmap

- 1 GPs and Differential Equations
- 2 Cascaded Differential Equations
- 3 Non-linear Response Models
- 4 Discussion and Future Work
- 5 Acknowledgements

- Integration of probabilistic inference with mechanistic models.
- These results are small simple systems.
- Ongoing work:
 - ▶ Scaling up to larger systems
 - ▶ Applications to other types of system, e.g. non-steady-state metabolomics, spatial systems etc.
 - ▶ Improved approximations.
 - ▶ Stochastic differential equations

Outline

- 1 GPs and Differential Equations
- 2 Cascaded Differential Equations
- 3 Non-linear Response Models
- 4 Discussion and Future Work
- 5 Acknowledgements

Acknowledgements

- Investigators: Neil Lawrence and Magnus Rattray
- Researchers: Peo Gao, Antti Honkela, Michalis Titsias and Jennifer Withers
- Charles Girardot and Eileen Furlong of EMBL in Heidelberg (mesoderm development in *D. Melanogaster*).
- Martino Barenco and Mike Hubank at the Institute of Child Health in UCL (p53 pathway).
- Raya Khanin and Ernst Wit of the University of Glasgow and the University of Lancaster (*E. coli* repressor system).

Funded by the BBSRC award “Improved Processing of microarray data using probabilistic models” and EPSRC award “Gaussian Processes for Systems Identification with applications in Systems Biology”

References I

- M. Barenco, D. Tomescu, D. Brewer, R. Callard, J. Stark, and M. Hubank. Ranked prediction of p53 targets using hidden variable dynamic modeling. *Genome Biology*, 7(3):R25, 2006. [PDF].
- R. T. Cirz, J. K. Chin, D. R. Andes, V. de Crécy-Lagard, W. A. Craig, and F. E. Romesberg. Inhibition of mutation and combating the evolution of antibiotic resistance. *PLoS Biology*, 3(6), 2005.
- J. Courcelle, A. Khodursky, B. Peter, P. O. Brown, , and P. C. Hanawalt. Comparative gene expression profiles following UV exposure in wild-type and SOS-deficient *Escherichia coli*. *Genetics*, 158:41–64, 2001.
- P. Gao, A. Honkela, M. Rattray, and N. D. Lawrence. Gaussian process modelling of latent chemical species: Applications to inferring transcription factor activities. *Bioinformatics*, 24:i70–i75, 2008. [PDF]. [DOI].
- R. Khanin, V. Viciotti, and E. Wit. Reconstructing repressor protein levels from expression of gene targets in *E. Coli*. *Proc. Natl. Acad. Sci. USA*, 103(49):18592–18596, 2006. [PDF]. [DOI].
- A. M. Lee, C. T. Ross, B.-B. Zeng, , and S. F. Singleton. A molecular target for suppression of the evolution of antibiotic resistance: Inhibition of the *Escherichia coli* RecA protein by N6-(1-Naphthyl)-ADP. *J. Med. Chem.*, 48(17), 2005.
- S. Rogers and M. Girolami. Model based identification of transcription factor regulatory activity via Markov chain Monte Carlo. Presentation at MASAMB '06, 2006.
- C. K. I. Williams. Computing with infinite networks. In M. C. Mozer, M. I. Jordan, and T. Petsche, editors, *Advances in Neural Information Processing Systems*, volume 9, Cambridge, MA, 1997. MIT Press.